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新型抗菌药物头孢地尔耐药现状及耐药机制研究进展*

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摘要: 头孢地尔是一种新型的儿茶酚-铁载体头孢菌素, 具有治疗多种细菌感染的潜力, 如铜绿假单胞菌、鲍曼不动杆菌、嗜麦芽窄食单胞菌感染等。尽管头孢地尔尚未在我国上市, 但已经有文献报道了对头孢地尔的耐药菌株。该文综述了头孢地尔的特点及抗菌作用、临床常见细菌对头孢地尔的耐药情况、转铁蛋白基因突变及 β -内酰胺酶、PBP 突变等方面的耐药研究现状。然而国内关于头孢地尔耐药机制的研究报道相对较少, 还需要做进一步的研究来更深入地了解头孢地尔的耐药机制。

关键词: 头孢地尔; 耐药; 转铁蛋白; β -内酰胺酶; PBP 突变**中图分类号:** R978.1 **文献标志码:** A **文章编号:** 1672-9455(2025)02-0285-04

Research progress on current situation and mechanisms of resistance of new antibiotic drug cefiderocol*

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Abstract: Cefiderocol is a novel catechol-iron carrier cephalosporin, which has the potential to treat a variety of bacterial infections, such as *Pseudomonas aeruginosa*, *Acinetobacter baumannii* and *Stenotrophomonas maltophilia*. Although the drug has not yet been marketed in China, there have been literature reports of drug-resistant strains of cefiderocol. This article reviews the characteristics and antimicrobial actions of cefiderocol, the resistance current situation of clinical common pathogens to it, transferrin gene mutations, β -lactamases and PBP mutations. However, there are relatively few reports in China on the resistance mechanisms of cefiderocol, so further research is needed to gain a deeper understanding of its resistance mechanisms.

Key words: cefiderocol; drug resistance; transferrin; β -lactamases; PBP mutation

抗菌药物在临床治疗中的广泛使用, 耐药性发生率和碳青霉烯类耐药肠杆菌(CRE)的数量也有所增加^[1], 治疗由 CRE 引起的感染依赖于替加环素、多黏菌素或新的 β -内酰胺/ β -内酰胺酶抑制剂联合用药^[2]。但这些抗菌药物作为最后的治疗方法, 也出现耐药^[3], 如头孢他啶/阿维巴坦(CAZ/AVI)对 B 类 β -内酰胺酶[例如新德里金属 β -内酰胺酶(NDM)]的活性有限^[4]。文献报道头孢地尔在体外对革兰阴性病原菌具有强效抗菌活性, 包括多重耐药的肠杆菌目和非发酵菌, 如铜绿假单胞菌、鲍曼不动杆菌、伯克霍尔德菌^[5-7]。在我国, 头孢地尔虽然未上市, 但已有文献报道其耐药菌株^[8-10]。本文综述了头孢地尔在国内外的耐药情况和耐药机制。

1 头孢地尔的特点及抗菌作用

头孢地尔是一种新型的儿茶酚-铁载体头孢菌素,

分别于 2019 年和 2020 年获得美国食品药品监督管理局(FDA)和欧洲药品管理局的批准。头孢地尔的化学结构^[11](图 1): C-7 位侧链上羧基丙氧基氨基, 该基团不被 β -内酰胺酶水解; 同时具有 C-3 侧链上的吡咯烷酮基团, 该基团能防止被 β -内酰胺酶识别。头孢地尔作用机制区别于头孢他啶和头孢吡肟在于邻苯二酚基团, 该结构能螯合三价铁离子, 然后依赖 TonB 系统的亲铁受体蛋白(TBDTs)进行转运。常见的 TBDTs 主要有 FepA、FecA、FhuA、CirA、Fiu、BtuB 和 FhuE, 主要负责运输邻苯二酚型亲铁素的有 FepA、CirA 和 Fiu^[12]。

天然铁载体根据与铁离子配位的功能基团被分为 4 种类型: 羟肟酸、邻苯二酚、羧基羧酸和混合型^[13], 头孢地尔的铁载体成分是邻苯二胺。头孢地尔上的邻苯二酚基团与铁载体结合, 如 CirA, 通过位于

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细菌外膜上的 TBDTs (如 CirA 受体通道) 被转运到细菌的外周质间隙, 还可通过细菌铁转运通道主动进入细菌周质空间, 从而作用于青霉素结合蛋白 (PBPs), 从而抑制细胞壁的合成, 发挥杀菌作用。与其他仅依赖于孔蛋白介导的被动扩散进入细菌的抗菌药物相比, 即使在孔蛋白缺失或突变的细菌中, 也不影响头孢地尔进入细胞周质达到高浓度聚集。

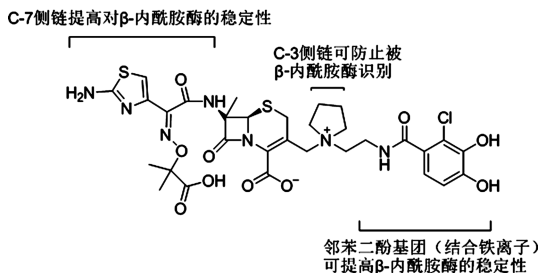


图 1 头孢地尔化学结构

2 临床常见细菌对头孢地尔的耐药情况

HACKEL 等^[14]的研究显示, 他们在 2014 年进行的大型国际监测中, 对头孢地尔的敏感性进行了测试。他们测试了 28 000 多株革兰阴性菌株, 包括一些碳青霉烯类耐药菌株, 如产生不同类型的 KPC 和 NDM 的菌株, 研究结果显示, 头孢地尔对这些菌株的活性保持不变。另外 HACKEL 等^[15]在 2014—2015 年进行的研究中, 纳入了从北美和欧洲分离的 9 205 株革兰阴性菌株, 包括肠杆菌目细菌、铜绿假单胞菌、鲍曼不动杆菌、嗜麦芽窄食单胞菌等, 其中有 68 株 (0.74%) 对头孢地尔不敏感。KARLOWSKY 等^[16]对北美和欧洲 2015—2016 年分离的 8 954 株革兰阴性菌株中, 包括肠杆菌目细菌、铜绿假单胞菌、鲍曼不动杆菌、嗜麦芽窄食单胞菌等, 有 51 株 (0.57%) 对头孢地尔具有耐药性。KAZMIERCZAK 等^[17]研究中, 纳入 1 272 株菌株, 包括 543 株产 OXA-23 酶、75 株产 KPC 酶、4 株产 IMP 酶、53 株产 VIM 酶、32 株产 OXA-48 酶、14 株产 NDM 酶、124 株产 OXA-24 酶、14 株 OXA-58 酶、420 株碳青霉烯类敏感菌株, 有 29 株 (2.28%) 对头孢地尔耐药, 其中 15 株产 OXA-23 酶、6 株产 OXA-24 酶、5 株产 NDM 酶, 此外还有 3 株碳青霉烯酶阴性分离株。WANG 等^[8]的研究显示, 1 158 株 CRE 分离株中, 碳青霉烯类耐药的肺炎克雷伯菌 (2 株) 和大肠埃希菌 (26 株) 对头孢地尔耐药, 26 株大肠埃希菌产 NDM-5 酶, 其中 1 株产 NDM-5 酶和 KPC-2 酶; 在耐碳青霉烯类大肠埃希菌 (181 株) 中, 有 26 株 (14.36%) 对头孢地尔具有耐药性, 且数据显示碳青霉烯类耐药大肠埃希菌中存在更多高水平的头孢地尔耐药分离株。LAN 等^[9]的研究中, 从 15 家三级医院的血液病患者血液中采集了 86 株非重复 CRKP 菌株, 有 4 株菌株对头孢地尔耐药, 4 株菌株均产生 NDM 酶。

3 头孢地尔耐药机制研究

3.1 转铁蛋白基因突变

大多数致病细菌在获取铁元素方面有多种途径, 参与铁运输的基因越多, 细菌

耐药的可能性就越大。头孢地尔可通过铁载体 CirA、FepA、IroN、FecA、FhuE、IutA、Fiu、Iha、FyuA 和 FitA 进入细菌内^[18]。当只敲除 CirA 基因时, 头孢地尔的最小抑菌浓度 (MIC) 没有变化, 而当 CirA 和 Fiu 双重基因敲除时, MIC 增加了 16 倍, 仍对头孢地尔敏感^[19], 这与 POIREL 等^[20]报道一致, CirA 基因的突变对头孢地尔的敏感性几乎没有影响。LAN 等^[21]对 CirA1 和 CirA198 两种等位基因进行研究, 在缺铁的情况下, CirA1 运输铁的能力比 CirA198 强, 当 CirA1 变异为 CirA198 时, MIC 提升 4 倍, 反之则降低 4 倍。KLEIN 等^[22]报道, 亲铁素受体 CirA 存在异质突变, 导致头孢地尔耐药。文献报道 Fiu 对具有邻二羟基苯甲酸基团或类似的二羟基吡啶基团的抗菌剂的敏感性也很重要^[23]。此外, 铜绿假单胞菌对头孢地尔的耐药与 piuD 和 pirR 基因相关, 鲍曼不动杆菌则与 pirA 基因和 piuA 基因有关^[24]。有研究报道, 细菌对头孢地尔耐药, 除了与产酶有关外, 与 pirA 基因编码的铁蛋白受体减少也有关^[25]。

3.2 β-内酰胺酶

在使用头孢他啶或头孢他啶/阿维巴坦时, KPC-2 酶 (D179A、D179Y、D179G) 和 OXA-48 (F72L、F156C/V) 突变体, 也会对头孢地尔产生交叉耐药, CMY-2 (S308R、S308N、D309G、L317P、S308N/D309G)、CTX-M-15 酶型则尚未发现对头孢地尔产生交叉耐药^[26-28]。KAZMIERCZAK 等^[17]研究中, 对头孢地尔耐药的菌株中, 有 15 株产 OXA-23 酶, 6 株产 OXA-24 酶, 5 株产 NDM 酶。WANG 等^[8]的研究中, 对头孢地尔耐药的 26 株大肠埃希菌产 NDM-5 酶, 其中 1 株产 NDM-5 酶和 KPC-2 酶。KOHIRA 等^[29]研究发现, 大肠埃希菌携带 NDM 和 PER 型 β-内酰胺酶等基因型菌株, 与头孢地尔的非敏感性有关。有文献报道, PER 基因和 NDM 基因导致鲍曼不动杆菌对头孢地尔的敏感性下降^[30-31]。β-内酰胺酶基因 bla_{SHV}-12 也被发现与肠杆菌科体外头孢地尔敏感性降低有关^[32]。

3.3 PBP 突变

PBPs 是细胞壁肽聚糖合成的必需酶, 也是 β-内酰胺类药物的重要靶点。2016 年, ITO 等^[19]报道测试了头孢地尔与大肠埃希菌不同 PBPs 的亲合力, 发现在大肠埃希菌的 PBPs 中, 头孢地尔对 PBP3 的亲合力最高。当在 PBP3 的 333 位插入序列 YRIN 或 YRIK^[8, 33], PBP3 发生突变时, 头孢地尔与靶点亲合力下降, 导致头孢地尔耐药^[25]。

3.4 其他耐药机制

此外, 细菌孔蛋白通道改变或丢失的突变, 如肺炎克雷伯菌中的 OmpK35 和 OmpK36, 与头孢地尔抗菌活性的轻微下降有关^[34]。预先接触某些头孢菌素、CZA 耐药 KPC 突变、AmpC 变体的出现等对头孢地尔耐药性有影响^[32]。

4 耐药性检测

目前美国临床和实验室标准化协会 (CLSI) 及欧洲抗微生物药物敏感性测试委员会 (EUCAST) 对头孢地尔药敏方法以及折点有指南共识, 肠杆菌目、铜

绿假单胞菌、鲍曼不动杆菌、嗜麦芽窄食单胞菌对头孢地尔具有明确的折点^[35]。然而,头孢地尔在与对碳青霉烯类耐药的病原体相互作用中存在广泛且未被察觉的异质耐药现象^[36];有研究利用纸片扩散法对头孢地尔进行敏感性测试时,当结果小于 17 mm 时,该方法有一定局限性,需要其他方法复核^[37]。

5 结 语

对头孢地尔的耐药机制主要包括转铁蛋白突变、 β -内酰胺酶、PBP 突变等,而 NDM 基因的表达和 CirA 基因缺陷的联合作用会导致高水平的头孢地尔耐药,单独因素会引起 MIC 微小变化。高水平头孢地尔耐药的主要原因是 CirA 失活时,头孢地尔进入的重要通道被关闭,进入细菌内的药物浓度急剧降低,使 NDM 酶充分水解头孢地尔,导致头孢地尔失去活性。

此外,国内关于头孢地尔的耐药机制的研究报道较少,是否存在异质耐药现象以及预先接触某些头孢菌素或者 CZA 耐药 KPC 变体或者 AmpC 变体的出现等导致头孢地尔耐药,均有待进一步研究。

参考文献

- [1] JERNIGAN J A, HATFIELD K M, WOLFORD H, et al. Multidrug-resistant bacterial infections in U. S. hospitalized patients, 2012 – 2017 [J]. *N Engl J Med*, 2020, 382 (14): 1309-1319.
- [2] RODRÍGUEZ-BAÑO J, GUTIÉRREZ-GUTIÉRREZ B, MACHUCA I, et al. Treatment of infections caused by extended-spectrum-beta-lactamase-, AmpC-, and carbapenemase-producing enterobacteriaceae [J]. *Clin Microbiol Rev*, 2018, 31(2): e00079-17.
- [3] LAXMINARAYAN R, VAN BOECKEL T, FROST I, et al. The Lancet Infectious Diseases Commission on antimicrobial resistance: 6 years later [J]. *Lancet Infect Dis*, 2020, 20(4): e51-e60.
- [4] SORIANO A, CARMELI Y, OMRANI A S, et al. Ceftazidime-avibactam for the treatment of serious gram-negative infections with limited treatment options: a systematic literature review [J]. *Infect Dis Ther*, 2021, 10(4): 1989-2034.
- [5] ABDUL-MUTAKABBIR J C, ALOSAIMY S, MORRIS-TE T, et al. Cefiderocol: a novel siderophore cephalosporin against multidrug-resistant gram-negative pathogens [J]. *Pharmacotherapy*, 2020, 40(12): 1228-1247.
- [6] BILAL M, EL TABELI L, BÜSKER S, et al. Clinical pharmacokinetics and pharmacodynamics of cefiderocol [J]. *Clin Pharmacokinet*, 2021, 60(12): 1495-1508.
- [7] JACOBS M R, ABDELHAMED A M, GOOD C E, et al. ARGONAUT- I: activity of cefiderocol (s-649266), a siderophore cephalosporin, against gram-negative bacteria, including carbapenem-resistant nonfermenters and enterobacteriaceae with defined extended-spectrum β -lactamases and carbapenemases [J]. *Antimicrob Agents Chemother*, 2019, 63(1): e01801-18.
- [8] WANG Q, JIN L, SUN S, et al. Occurrence of high levels

of cefiderocol resistance in carbapenem-resistant *escherichia coli* before its approval in China: a report from China CRE-network [J]. *Microbiol Spectr*, 2022, 10(3): e0267021.

- [9] LAN P, LU Y, CHEN Z, et al. Emergence of high-level cefiderocol resistance in carbapenem-resistant *klebsiella pneumoniae* from bloodstream infections in patients with hematologic malignancies in China [J]. *Microbiol Spectr*, 2022, 10(2): e0008422.
- [10] WANG C, YANG D, WANG Y, et al. Cefiderocol for the treatment of multidrug-resistant gram-negative bacteria: a systematic review of currently available evidence [J]. *Front Pharmacol*, 2022, 13: 896971.
- [11] MCCREARY E K, HEIL E L, TAMMA P D. New perspectives on antimicrobial agents: cefiderocol [J]. *Antimicrob Agents Chemother*, 2021, 65(8): e0217120.
- [12] PINKERT L, LAI Y H, PEUKERT C, et al. Antibiotic conjugates with an artificial MECAM-based siderophore are potent agents against gram-positive and gram-negative bacterial pathogens [J]. *J Med Chem*, 2021, 64(20): 15440-15460.
- [13] KRAMER J, OEZKAYA O, KUEMMERLI R. Bacterial siderophores in community and host interactions [J]. *Nat Rev Microbiol*, 2020, 18(3): 152-163.
- [14] HACKEL M A, TSUJI M, YAMANO Y, et al. In vitro activity of the siderophore cephalosporin, cefiderocol, against carbapenem-nonsusceptible and multidrug-resistant isolates of gram-negative bacilli collected worldwide in 2014 to 2016 [J]. *Antimicrob Agents Chemother*, 2018, 62(2): e01968-17.
- [15] HACKEL M A, TSUJI M, YAMANO Y, et al. In vitro activity of the siderophore cephalosporin, cefiderocol, against a recent collection of clinically relevant gram-negative bacilli from North America and Europe, including carbapenem-nonsusceptible isolates (SIDERO-WT-2014 Study) [J]. *Antimicrob Agents Chemother*, 2017, 61(9): e00093-17.
- [16] KARLOWSKY J A, HACKEL M A, TSUJI M, et al. In vitro activity of cefiderocol, a siderophore cephalosporin, against gram-negative bacilli isolated by clinical laboratories in north america and europe in 2015 – 2016; SIDERO-WT-2015 [J]. *Int J Antimicrob Agents*, 2019, 53(4): 456-466.
- [17] KAZMIERCZAK K M, TSUJI M, WISE M G, et al. In vitro activity of cefiderocol, a siderophore cephalosporin, against a recent collection of clinically relevant carbapenem-non-susceptible Gram-negative bacilli, including serine carbapenemase- and metallo- β -lactamase-producing isolates (SIDERO-WT-2014 Study) [J]. *Int J Antimicrob Agents*, 2019, 53(2): 177-184.
- [18] LI L L, YORIK O. Confined mobility of TonB and FepA in *escherichia coli* membranes [J]. *PLoS One*, 2016, 11(12): e0160862.
- [19] ITO A, NISHIKAWA T, MATSUMOTO S, et al. Siderophore cephalosporin cefiderocol utilizes ferric iron transporter systems for antibacterial activity against

- Pseudomonas aeruginosa* [J]. *Antimicrob Agents Chemother*, 2016, 60(12): 7396-7401.
- [20] POIREL L, SADEK M, NORDMANN P. Contribution of PER-type and NDM-type β -lactamases to cefiderocol resistance in *Acinetobacter baumannii* [J]. *Antimicrob Agents Chemother*, 2021, 65(10): e0087721.
- [21] LAN P, LU Y, JIANG Y, et al. Catecholate siderophore receptor CirA impacts cefiderocol susceptibility in *Klebsiella pneumoniae* [J]. *Int J Antimicrob Agents*, 2022, 60(4): 106646.
- [22] KLEIN S, BOUTIN S, KOCER K, et al. Rapid development of cefiderocol resistance in carbapenem-resistant enterobacter cloacae during therapy is associated with heterogeneous mutations in the catecholate siderophore receptor cirA [J]. *Clin Infect Dis*, 2022, 74(5): 905-908.
- [23] GRINTER R, LITHGOW T. The structure of the bacterial iron-catecholate transporter Fiu suggests that it imports substrates via a two-step mechanism [J]. *J Biol Chem*, 2019, 294(51): 19523-19534.
- [24] KARAKONSTANTIS S, ROUSAKI M, KRITSOTAKIS E I. Cef-iderocol: systematic review of mechanisms of resistance, heteroresistance and in vivo emergence of resistance [J]. *Antibiotics (Basel)*, 2022, 11(6): 723.
- [25] MALIK S, KAMINSKI M, LANDMAN D, et al. Cefiderocol resistance in *Acinetobacter baumannii*: roles of β -lactamases, siderophore receptors, and penicillin binding protein 3 [J]. *Antimicrob Agents Chemother*, 2020, 64(11): e01221-20.
- [26] HOBSON C A, COINTE A, JACQUIER H, et al. Cross-resistance to cefiderocol and ceftazidime-avibactam in KPC β -lactamase mutants and the inoculum effect [J]. *Clin Microbiol Infect*, 2021, 27(8): 1172. e7-1172. e10.
- [27] BIANCO G, BOATTINI M, COMINI S, et al. In vitro activity of cefiderocol against ceftazidime-avibactam susceptible and resistant KPC-producing Enterobacterales: cross-resistance and synergistic effects [J]. *Eur J Clin Microbiol Infect Dis*, 2022, 41(1): 63-70.
- [28] FRÖHLICH C, SØRUM V, TOKURIKI N, et al. Evolution of β -lactamase-mediated cefiderocol resistance [J]. *J Antimicrob Chemother*, 2022, 77(9): 2429-2436.
- [29] KOHIRA N, WEST J, ITO A, et al. In vitro antimicrobial activity of a siderophore cephalosporin, S-649266, against enterobacteriaceae clinical isolates, including carbapenem-resistant strains [J]. *Antimicrob Agents Chemother*, 2016, 60(2): 729-734.
- [30] POIREL L, KIEFFER N, NORDMANN P. Stability of cefiderocol against clinically significant broad-spectrum oxacillinases [J]. *Int J Antimicrob Agents*, 2018, 52(6): 866-867.
- [31] LIU X, LEI T, YANG Y, et al. Structural basis of PER-1-mediated cefiderocol resistance and synergistic inhibition of PER-1 by cefiderocol in combination with avibactam or durlobactam in *Acinetobacter baumannii* [J]. *Antimicrob Agents Chemother*, 2022, 66(12): e0082822.
- [32] LIU C, YI J, LU M, et al. Dynamic within-host cefiderocol heteroresistance caused by bla_{SHV}-12 amplification in pandrug-resistant and hypervirulent *Klebsiella pneumoniae* sequence type 11 [J]. *Drug resistance updates*, 2024, 73: 101038.
- [33] ALM R A, JOHNSTONE M R, LAHIRI S D. Characterization of *Escherichia coli* NDM isolates with decreased susceptibility to aztreonam/avibactam: role of a novel insertion in PBP3 [J]. *J Antimicrob Chemother*, 2015, 70(5): 1420-1428.
- [34] SATO T, YAMAWAKI K. Cefiderocol: discovery, chemistry, and in vivo profiles of a novel siderophore cephalosporin [J]. *Clin Infect Dis*, 2019, 69(Suppl 7): S538-S543.
- [35] GOLDEN A R, ADAM H J, BAXTER M, et al. In vitro activity of cefiderocol, a novel siderophore cephalosporin, against gram-negative bacilli isolated from patients in canadian intensive care units [J]. *Diagn Microbiol Infect Dis*, 2020, 97(1): 115012.
- [36] CHOBY J E, OZTURK T, SATOLA S W, et al. Widespread cefiderocol heteroresistance in carbapenem-resistant gram-negative pathogens [J]. *Lancet Infect Dis*, 2021, 21(5): 597-598.
- [37] BIANCO G, BOATTINI M, COMINI S, et al. Disc Diffusion and ComASP[®] cefiderocol microdilution panel to overcome the challenge of cefiderocol susceptibility testing in clinical laboratory routine [J]. *Antibiotics (Basel)*, 2023, 12(3): 604.

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(上接第 284 页)

- [36] 万诗扬. 补体 C1q、COP-LMR 评分等外周血指标与非小细胞肺癌免疫治疗预后相关性研究 [D]. 大连: 大连医科大学, 2022.
- [37] KAIRA K, HIGUCHI T, NARUSE I, et al. Metabolic activity by ¹⁸F-FDG-PET/CT is predictive of early response after nivolumab in previously treated NSCLC [J]. *Eur J Nucl Med Mol Imaging*, 2018, 45(1): 56-66.
- [38] WANG Y, ZHAO N, WU Z B, et al. New insight on the correlation of metabolic status on ¹⁸F-FDG PET/CT with immune marker expression in patients with non-small cell lung cancer [J]. *Eur J Nucl Med Mol Imaging*, 2020, 47(5): 1127-1136.
- [39] MONACO L, GEMELLI M, GOTUZZO I, et al. Metabolic parameters as biomarkers of response to immunotherapy and prognosis in Non-Small cell lung cancer (NSCLC): a real world experience [J]. *Cancers (Basel)*, 2021, 13(7): 1634.
- [40] 尚士洁. 晚期非小细胞肺癌患者血脂水平与免疫治疗疗效的相关性研究 [D]. 济南: 山东大学, 2023.
- [41] 张璐锦. 糖脂代谢与非小细胞肺癌免疫检查点抑制剂疗效关系的研究 [D]. 开封: 河南大学, 2023.

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